

Enhanced Enzyme Inhibitory Effects of the Nanohybrid Eggplant Extract: An Unusual Pharmaceutical Form for Medicinal Plant

Ceylan Dönmez¹, Ufuk Koca-Çalışkan^{2,3}, Nuraniye Eruygur¹, Cevahir Altınkaynak⁴, Nalan Özdemir⁵

¹ Selçuk University, Faculty of Pharmacy, Department of Pharmacognosy, Konya, Türkiye.

² Gazi University, Faculty of Pharmacy, Department of Pharmacognosy, Ankara, Türkiye.

³ Düzce University, Faculty of Pharmacy, Department of Pharmacognosy, Düzce, Türkiye.

⁴ Nevşehir Hacı Bektaş Veli University, Avanos Vocational School, Department of Plant and Animal Production, Nevşehir, Türkiye.

⁵ Erciyes University, Faculty of Science, Chemistry Department, Kayseri, Türkiye.

Correspondence Author: Ceylan Dönmez

E-mail: ceylan.donmez@selcuk.edu.tr

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ABSTRACT

Objective: Recently, biosynthesis/synthesis of nanoflowers has become very attractive for chemical and pharmaceutical sciences, and enhanced enzyme activities. Various plant extracts and their active compounds are effectively used as organic component for novel nanoflowers synthesis. *Solanum melongena* L., commonly known as eggplant in English, a vegetable and medicinal plant belongs to Solanaceae family has several advantages in materials synthesis due to cheap and obtained easily. The aim of this study is to compare the enzyme ((alpha-glucosidase (AGase), alpha-amylase (AAase), tyrosinase (Tyr), acetylcholinesterase (AChE) and butyryl cholinesterase (BChE)) inhibitory effects of the eggplant' calyx extract and its Solanum-inorganic hybrid nanoflower (Sm-ihNFs) via *in vitro* experimental methods.

Methods: The hybrid nanoflower was formed (NF) with organic molecules, eggplant extract (Sm), and inorganic compounds, copper to enhance the catalytic activities. The inhibition capacities of the eggplant extract, and its hybrid nanoflower were evaluated on selected enzymes (AGase, AAase, Tyr, AChE and BChE) which play significant roles physiologically by *in vitro* tests in this study.

Results: According to inhibition percentages and IC₅₀ values, Sm-ihNFs showed higher inhibitory activities on enzymes other than ache than the plain crude plant extract. Among all the enzymes that were studied, Sm-ihNFs demonstrated significantly higher alpha-glucosidase and alpha-amylase inhibition activities compared to acarbose. And when compared to galantamine hydrobromide Sm-ihNFs showed higher enzyme inhibition and significant IC₅₀ value.

Conclusion: It was thought that Sm-ihNFs prepared from eggplant extract may have promising potential for antidiabetic drug formulations in the future. The hybrid nanoflowers will be promising and guide for the future work in terms of pharmaceutical and cosmeceutical industry.

Keywords: *Solanum melongena*; nanoflower; alpha-glucosidase amylase; tyrosinase; cholinesterases

1. INTRODUCTION

Solanum melongena L. (Solanaceae), commonly known as eggplant, is one of the most consumed, economically valuable vegetables in most of the world (1). The aerial parts of the plant are rich in alkaloids and saponins, phenolics (2,3). Dönmez et al. examined whether methanolic eggplant extract has antihemorrhoidal and antioxidant activities on rats or not. They have used eggplant calyx for preparing the extract. As a result of the study, it has been shown that the extract has high biological activity (3). Medicinal features such as analgesic, antifungal, antiasthmatic, antidiabetic, antihemorrhoidal, antiinflammatory, antipyretic, antioxidant, antiplatelet, hypocholesterolemic, hypolipidemic, hypotensive, and spasmogenic activities of the plant have been scientifically proven (4-19). Umamageswari and Maniyar demonstrated that eggplant leaf extract has antiinflammatory activity, which

is related to many enzymes (20). Previously, Kwon et al. had evaluated just α -amylase (AAase), α -glucosidase (AGase) and angiotensin I-converting enzyme (ACE) inhibitory activities of eggplant extract to investigate a potential drug for type 2 diabetes and hypertension (21).

In recent years, innovative ideas on the development of biomaterials using herbal extracts have been putting forward and many plant-derived nanostructures and nanoparticles are tried to be developed with different methods (22). Hybrid nanoflower is a novel nano-bio agent formed with organic and inorganic compounds, moreover, creating hybrid nanoflowers using plant extracts and their constituents, which is green chemistry, became attractive yet convenient (23-29). It has been reported so far that nanoflowers obtained

using plant extract exhibit improved biological activity. The organic-inorganic hybrid nanoflower (TF-Cu²⁺ hNF) was obtained using *Trigonella foenum-graecum* L. extracts with copper ions. The TF-Cu²⁺ hNF exhibited enhanced antibacterial activity against *Enterococcus faecium*, *E. faecalis*, *Staphylococcus aureus*, *Bacillus cereus*, *Salmonella typhi* and *Escherichia coli* except against *Pseudomonas aeruginosa* and *Haemophilus influenza* (30). The TF-Cu²⁺ hNFs and free TF extracts were also reported to show no antifungal activity at the same research. The synthesis of *Viburnum opulus* (European cranberry bush) extract based Cu²⁺ hybrid nano forms called “snowballs” and their effective catalytic and antimicrobial features compared to free European cranberry bush extracts were reported (31). They suggested that biologic activities may follow a fenton-like pathway. Baldemir and coworkers reported the synthesis and characterization method for hybrid nanoflower with using green tea extract and copper ion. Baldemir and coworkers reported the synthesis and characterization method for hybrid nanoflower with using *Camellia sinensis* (L.) Kuntze extract and copper ion (32). They also demonstrated enhanced antimicrobial and catalytic activities of nanoflowers. In the last nanoflower study using curcumin extract, the effect of reaction time and curcumin concentrations on nanoflower’s morphology was reported (33).

Solanum melongena plant extracts with their rich alkaloid, saponin and phenolic constituents can be used in hybrid nanoflower synthesis. However, no study was recorded on enzymes inhibitory activities of calyx and the nanoflower form of the extract to this point. In addition to antidiabetic enzyme activities, it was aimed to evaluate anticholinesterase and tyrosinase enzyme inhibition activities in this article. In this study, which allows comparison of antidiabetic activity, the anticholinesterase and tyrosinase enzyme inhibition activities of calyxes were examined for the first time. For this; the hybrid nanoflowers using eggplant extract were obtained in optimum conditions, then characterized with several techniques such as Scanning Electron Microscope (SEM), Energy Dispersive X-Ray Analysis (EDX), Fourier Transform Infrared (FTIR) and X-Ray Diffraction (XRD). Then, some important enzyme inhibition activities such as alpha-glucosidase (AGase), alpha-amylase (AAase), tyrosinase (Tyr), acetylcholinesterase (AChE) and butyryl cholinesterase (BChE) of hybrid nanoflowers were investigated in comparison with the free extract. We suggested that enhanced enzymatic activities of nanoflowers may expand their uses a usual pharmaceutical form for an edible/medicinal plant.

2. METHODS

2.1. Preparation of the Extract

The cultured plant materials, which was founded in the Herbarium of Faculty of Pharmacy/Gazi University/Ankara/Turkey as authenticated voucher specimen (GUE 3500), were gathered during August-September 2016-2017 at the Central

Anatolia of Turkey. Only the dark-green calyx of the mature eggplant fruit was separated and dried in shade, then ground to powder, extracted with methanol by maceration. The yield of the extract was calculated as 11.4 percent.

2.2. Synthesis of Hybrid Nanoflowers (Sm-ihnfs)

Sm-ihNFs was synthesized following modified reported method (31, 34). The calyx methanolic extract with concentrations from 0.02 to 0.2 mg mL⁻¹ was separately added into the mixture containing 50 mL of 10 mM PBS (pH7.4) and 0.8 mM Cu²⁺ ion. The occurring blueish-green precipitates, washed with water, and centrifuged at 5000 rpm for 10 min. The washing process was repeated 3 times and the final product was dried at 50°C for further characterizations and enzyme assays. Sm-ihNFs was characterized by using the morphologies of the synthesized Sm-ihNFs were examined by using Scanning Electron Microscopy (ZEISS EVO-LS10). The chemical and crystal structures of the Sm-ihNFs were characterized using Fourier Transform Infrared spectroscopy (Perkin Elmer 400, Spectrometer Spotlight 400 Imaging System) and X-Ray Diffraction (Bruker, AXS D8 Advance Model) analysis, respectively. The elemental analysis of the Sm-ihNFs was performed by energy dispersive X-ray (ZEISS EVO-LS10) analysis.

2.3. Enzyme inhibition activity

Alpha-glucosidase inhibition (35): The inhibition method of α -glucosidase was followed according to Kumar et al. Acarbose was utilized as reference. The test solution (25 μ L) was diluted with a PBS which added to α -glucosidase (25 μ L, 0.5 U/mL). After the 10 minutes incubation at 25°C, 25 μ L of 0.5 mM PNPG was added to each well then, the mixture was further waited for 30 minutes at 37°C. At the end of the incubation term, sodium carbonate (0.2 M, 100 μ L) was joined to put an end to the reaction and the absorbances were read at 405 nm. All concentrations were carried out in triplicate to obtain an accurate statistical analysis.

Alpha-amylase inhibition (36): The inhibition method of α -alpha-amylase was followed by Kumar et al. While acarbose was a positive control, PBS (pH 6.9, 0.02 M, PBS) was a negative control in place of the specimen. Each specimen was conducted in triplicate with diverse concentrations. The reaction mix containing 50 μ L of test solution was diluted with buffer, 25 μ L of enzyme (5000 μ g/mL, α -amylase) and incubated for about 10 minutes at 25°C. Then 50 μ L of freshly prepared 0.5 % starch solution (w/v) was annexed to each well as a substrate and incubated for a further 10 minutes at 25°C. Incubation period was followed by addition of 1 % 3,5-dinitrosalicylic acid (DNS, 100 μ L) coloring reagent and heated in a water bath for 10 minutes. The absorbances were read at 540 nm.

Tyrosinase enzyme inhibition (37): Tyrosinase inhibition was detected by the improved dopa chrome method. L-DOPA was used as substrate. A part of the plant extracts and their nanoflowers, under the optimal conditions, diffused in DMSO

(PBS, 6.8 pH, 80 μ L), 40 μ L of L-DOPA, and 40 μ L tyrosinase enzymes were added into each well. Analysis was performed in a microplate by using ELISA plate reader and absorbances were measured at 475 nm. Results were evaluated by comparing with the DMSO and α -kojic acid.

The AChE and BChE inhibition (38,39): The acetylcholinesterase/butyryl cholinesterase inhibition assay was evaluated by Ellman colorimetric method as described by Öztürk. 150 μ L of 0.1 M PBS (pH=8.0), 10 μ L of test solutions in MeOH: DMSO (4k: 1k, v/v) with different concentrations and 20 μ L of 0.1 U/mL enzyme solution (acetylcholinesterase-butyryl cholinesterase was obtained from equine serum) were incubate for 15 minutes at 25°C. 10 μ L of a solution of 0.2 mM (acetylcholine/butyryl thiocholine) and 10 μ L of 0.5 mM DTNB were mixed and the absorbances of the mix were measured at 412 nm. Galantamine hydrobromide (Sigma-Aldrich, Germany) was used as a positive control.

2.4. Statistical analysis

The differences in values between the reference and test groups were compared by using GraphPad Prism Software (8.3 Version, La Jolla/CA/USA) in statistical analysis. The results are expressed as the mean \pm standard error means (S.E.M.). When one-way ANOVA with Tukey Multiple Comparison Test was used, p-values of less than 0.05 were considered statistically significant.

3. RESULTS

3.1. Synthesis of Hybrid Nanoflowers (Sm-Ihnf)

Sm-ihNFs was synthesized using calyx extract of the plant in phosphate buffered saline and Cu (II) ion for 3 days of incubation. The extract contains diverse types of phytochemicals, such as alkaloids, saponosides, flavonoids that comprise important elements such as N, O and S atoms, which can form complexes with Cu (II) ions due to their strong affinity. In the synthesis of Sm-ihNFs, the most significant interaction is the coordination chemistry between Cu (II) ions and some molecules, mostly alkaloids containing N atom, which allows the formation of the hybrid structures with flower-like shapes under definite circumstances.

Formation of hybrid nanoflower consists of three consecutive steps as nucleation, growth, and completion. Nucleation occurs formation of the key nanocrystals of copper phosphate from Cu (II) and phosphate ions interactions. The reaction with primary nanocrystals and the biomolecules leads to the formation of the petals of flowers-like shapes during growth. As a result of penetration of petal-like structures, hybrid nanoflower formation occurs.

Concentration of organic component is one of the most essential parameters that influence the morphology and the structure of organic-inorganic hybrid nanoflowers. Consequently, effect of the concentrations of the extract on the morphology of Sm-ihNFs was investigated applying SEM

analysis (Fig.1). The most uniform morphology of hybrid nanoflowers was obtained as the optimum condition. The average size of nanoflowers was 5-6 μ m.

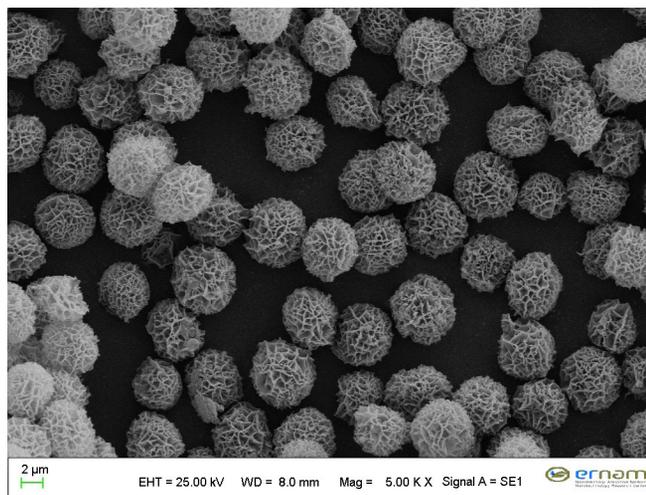


Figure 1. Scanning electron microscopic images of hybrid nanoflowers formed with 0.1 mg mL⁻¹ extract of *S. melongena*

The elemental analysis of the synthesized Sm-ihNFs was performed by EDX (Fig.2). The EDX spectrum was revealed presence of major elements such as C, N, O, P, and Cu in the structure. The copper metal can be evaluated as a major skeleton of the hybrid nanoflower.

The functional groups in the structure of organic compounds, whether the two compounds are the same, the state of the bonds in the structure, the binding sites and whether the structure is aromatic or aliphatic were determined with the FTIR analysis, (Fig.3). The bending and stretching bonds in the free plant extract were characteristically also detected in the hybrid structure.

The measurement of particle sizes, phase equilibria and crystal structure of Sm-ihNFs were investigated by XRD (Fig.4). The all diffraction peaks of Cu₃(PO₄)₂·3H₂O were matched well with the JCPDS card (022-0548).

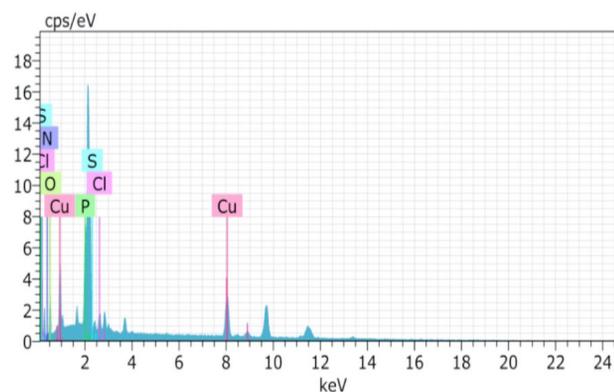


Figure 2. EDX spectra of hybrid nanoflowers formed with 0.1 mg mL⁻¹ extract of *S. melongena*

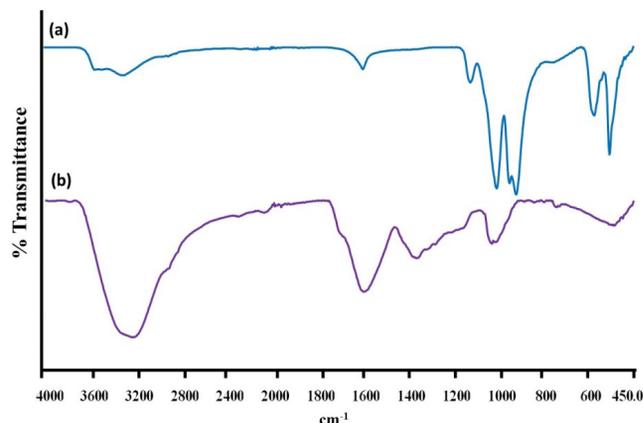


Figure 3. FT-IR spectrums of (a) hybrid nanoflowers formed with 0.1 mg mL⁻¹ extract of *S. melongena* and (b) the plant extract

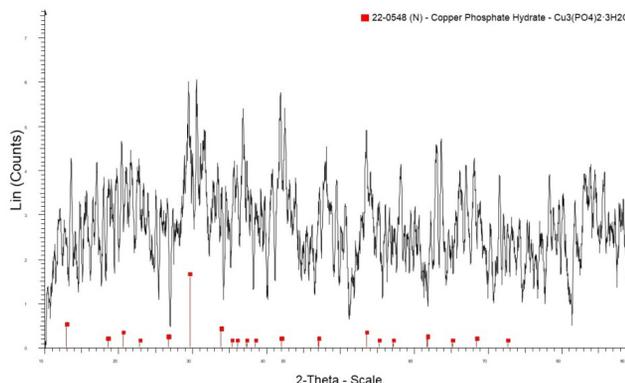


Figure 4. XRD spectra of hybrid nanoflowers formed with 0.1 mg mL⁻¹ extract of *S. melongena*

Table 1. Anticholinesterase activity (Inhibition % ± S.E.M. & IC₅₀ value) of eggplant extract and Sm-ihNFs

Samples	Anticholinesterase activity			
	AChE		BChE	
	Inhibition % #	IC ₅₀ (ppm)	Inhibition % #	IC ₅₀ (ppm)
Sm	70.12 ± 0.15***	17.09 ± 1.36**	75.76 ± 0.65**	28.29 ± 4.30**
Sm-ihNFs	13.83 ± 1.93***	321.46 ± 6.31***	85.12 ± 1.87*	32.76 ± 8.90**
Galantamine hydrobromide	92.23 ± 0.65***	2.31 ± 0.19**	81.82 ± 0.56*	3.01 ± 0.22*

Inhibition % ± S.E.M. at 0.2 mg mL⁻¹ concentrations

*p<.5, **p<.05, ***p<.005, ANOVA test

Table 2. Antidiabetic activity (Inhibition % ± S.E.M. & IC₅₀ value) of eggplant extract and Sm-ihNFs

Samples	Antidiabetic activity			
	α-glucosidase		α-amylase	
	Inhibition % #	IC ₅₀ (ppm)	Inhibition % #	IC ₅₀ (ppm)
Sm	46.67 ± 4.64*	212.33 ± 17.19***	42.05 ± 5.42**	213.80 ± 21.68***
Sm-ihNFs	82.77 ± 1.31***	31.95 ± 6.72**	83.06 ± 2.92***	27.81 ± 15.21**
Acarbose	57.74 ± 0.83*	59.27 ± 15.09*	58.40 ± 0.61*	38.46 ± 11.73**

Inhibition % ± S.E.M. at 0.2 mg mL⁻¹ concentrations

*p<.5, **p<.05, ***p<.005, ANOVA test

Table 3. Tyrosinase enzyme inhibition (Inhibition % ± S.E.M. & IC₅₀ value) of eggplant extract and Sm-ihNFs

Samples	Tyrosinase inhibition activity	
	Inhibition % #	IC ₅₀ (ppm)
Sm	4.59 ± 3.53*	704.56 ± 39.23***
Sm-ihNFs	16.43 ± 0.31**	423.80 ± 2.02**
Kojic acid	24.23 ± 5.52**	371.80 ± 36.80**

Inhibition % ± S.E.M. at 0.2 mg mL⁻¹ concentrations

*p<.5, **p<.05, ***p<.005, ANOVA test

3.2. Enzyme inhibitions

According to average cholinesterases inhibition ± standard error means (S.E.M.) values were given in Table 1, the BChE activity of nanoflower was higher than the galantamine

hydrobromide, whereas the AChE inhibition activity was higher in the plain extract.

According to average of the α-glucosidase and α-amylase inhibition ± standard error mean (S.E.M.) inhibition values

were given in Table 2, the enzyme inhibition of the samples was elevated compared to the reference acarbose.

Average Tyr inhibition \pm standard error means (S.E.M.) were demonstrated by the calculation of absorbance of the plain extract and its hybrid nanoflowers against blank sample. According to inhibition values given in Table 3, the results showed that the enzyme inhibition of the samples was low compared to kojic acid.

4. DISCUSSION

The prevalence of chronic and metabolic disorders continues to increase in humans around the world. The tendency to herbal resources has risen for the purpose of prevention or treatment of diseases (40). Today, many scientists have focused on the integration of natural resources and technological developments. In our study, nanoflower-plant hybridization studies were carried out to increase the biological activity potential of plants. One of the most important reasons for using *Solanum melongena* (eggplant) in this study was limited pharmacological studies on it despite its economically valuable. The present study demonstrated the green synthesis, characterization, and activity of the organic-inorganic hybrid Sm-ihNFs enzymes inhibitors. This easy to apply and one-step immobilization withstand the relation reactions between amine groups in the character strength of enzymes and cupric phosphate nanoflowers. Three consecutive steps were conducted for forming the nanoflower-like structure. First step was nucleation and formation of primary nano crystals. Second step was arranging crystals, and last step was formation of nanoflowers (23).

The results were promising that the hybrid nanostructures were synthesized demonstrated appealing blooming structures as 5-6 μm size. Likewise, characterization of the Sm-ihNFs was successfully completed by using different techniques (SEM, EDX, FTIR, and XRD). In a period of approximately 30-35 years, pharmacological and phytochemical studies on *Solanum species* have gained importance. At least 65 species (including *S. aculeastrum*, *S. aethiopicum*, *S. americanum*, *S. anguivi*, *S. cathayanum*, *S. capsicoides*, *S. diphyllum*, *S. muricatum*, *S. nigrum*, *S. septemlobum*, *S. sessiliflorum*, *S. spirale*, *S. surattense*, *S. torvum*, *S. tuberosum*, *S. violaceum*, and *S. xanthocarpum*) have been studied on scientific platform. The most antioxidants and anticancer activities of *Solanum species* have been demonstrated by scientists. The responsible compounds of pharmacological activities, such as antibacterial, anticancer, anticonvulsant, antidiabetic, anti-fungal, anti-inflammatory, antileishmanial, antioxidants, antitumor, and spasmolytic, of *Solanum species* have been attributed to steroidal alkaloids (41-43). There have been some publications related to the subject of this manuscript despite the small number of pharmacological studies on eggplant. One of these studies was conducted by Kwon and coworkers. They indicated that eggplants have radical scavenging-linked antioxidant activity and α -glucosidase inhibitory effect due to rich

phenolic compounds (10). That result supported our study, moreover, the hybrid nanoflower exhibited higher effective α -glucosidase and α -amylase enzymes activities than the plain plant extract. In another *in vitro* study, calystegines, a tropane alkaloid derivative in eggplant, showed glycosidases inhibitory activity (44). Consequently, the hybrid nano-flower might contain the N including alkaloid structure, therefore.

In a poster presentation presented by Ketprayoon and Chaicharoenpong, while *S. melongena*'s methanolic fruit extract showed low tyrosinase inhibitor activity, tyrosinase inhibitor activity of *S. melongena*'s aqueous fruit extract could not be determined (45). Although no significant activity was observed in both the calyx extracts and the hybrid nanoflower, higher effective tyrosinase inhibitor activity was determined in the hybrid nanoflower in our study. In another study, the ethanolic eggplant peels extract have showed the skin protecting (46). There is a need for evaluation of different eggplant parts and their various extracts.

The cholinesterase inhibitors are still the most important clinical strategy to manage Alzheimer's disease. In this study, while Sm-ihNF showed BChE enzyme inhibitory effect, it was not very successful on AChE enzyme inhibition. However, this is, to the best of our knowledge, the first report of the analysis of both acetyl and butyryl choline esterase enzymes inhibition activities of eggplant. It is thought that *Solanum* glycoalkaloids may be the compounds responsible for anticholinesterase activities (47).

5. CONCLUSION

The enzyme inhibition tests have been used for getting an idea about several different illness due to their activity and specificity performance. Although there are many enzyme inhibition studies on plants, studies on nano technological-chemical studies for increasing activity based on natural sources are limited. With the studies shown in this text, a novel and elegant immobilization approach was used to form nanoflower-like structures from enzymes exhibiting highly enhanced catalytical activity and stability. The nanoflower's large surface area, limited the mass-transfer, and nanoscale-related enzyme cooperation are potential causes in enzyme inhibition tests and biological activity studies. The synthesis of protein-inorganic hybrid nanoflower containing plant extracts is an important factor considering its suitability for commercial applications in pharmacy and cosmetics.

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REFERENCES

- [1] Van Eck J, Snyder A. Eggplant (*Solanum melongena* L.). *Methods Mol. Biol.* 2006; 343: 439-47. DOI: 10.1385/1-59745-130-4:439
- [2] W.C. Evans, Trease and Evans Pharmacognosy, 15th ed. Sanders Co. Ltd.: Singapore; 2002: 33-35.
- [3] Dönmez C, Yalçın FN, Boyacıoğlu Ö, Korkusuz P, Akkol EK, Nemutlu E, Balaban HY, Çalışkan, UK. From nutrition to medicine: Assessing hemorrhoid healing activity of *Solanum melongena* L. via *in vivo* experimental models and its major chemicals. *J. Ethnopharmacol.* 2020; 261; 113-143. DOI: 10.1016/j.jep.2020.113143
- [4] Vohora SB, Kumar I, Khan MSY. Effect of alkaloids of *Solanum melongena* on the central nervous system. *J. Ethnopharmacol.* 1984; 11: 331-336. DOI: 10.1016/0378-8741(84)90078-3
- [5] Das J, Lahan, JP, Srivastava RB. *Solanum melongena*: A potential source of antifungal agent. *Indian J. Microbiol.* 2010; 50: 62-69. DOI: 10.1007/s12088.010.0004-2
- [6] Han SW, Tae J, Kim JA, Kim DK, Seo GS, Yun KJ, Lee YM. The aqueous extract of *Solanum melongena* inhibits PAR2 agonist-induced inflammation. *Clin Chim Acta* 2003; 328: 39-44. DOI: 10.1016/s0009-8981(02)00377-7
- [7] Bello SO, Muhammad B, Gammaniel KS, Aguye AI, Ahmed H, Njoku CH. Randomized double blind placebo controlled clinical trial of *Solanum melongena* L. fruit in moderate to severe asthmatics. *J. Med. Sci.* 2004; 4: 263-269. DOI: 10.3923/jms.2004.263.269
- [8] Bello SO, Muhammad BY, Gammaniel KS, Abdu-Aguye I, Ahmed H, Njoku CH, Salka AM. Preliminary evaluation of the toxicity and some pharmacological properties of the aqueous crude extract of *Solanum melongena*. *Res. J. Agric. Biol. Sci.* 2005; 1: 1-9.
- [9] Caliskan UK, Aka C, Oz MG. Plants Used in Anatolian Traditional Medicine for the Treatment of Hemorrhoid. *Rec. Nat. Prod.* 2017; 11: 235-250.
- [10] Kwon YI, Apostolidis E, Shetty K. *In vitro* studies of eggplant (*Solanum melongena*) phenolics as inhibitors of key enzymes relevant for type 2 diabetes and hypertension. *Bioresour. Technol.* 2008; 99: 2981-2988. DOI: 10.1016/j.biortech.2007.06.035
- [11] Mutalik S, Paridhavi K, Rao CM, Udupa N. Antipyretic and analgesic effect of leaves of *Solanum melongena* Linn. in rodents. *Indian J. Pharmacol.* 2003; 35: 312-315.
- [12] Nisha P, Nazar PA, Jayamurthy P. A comparative study on antioxidant activities of different varieties of *Solanum melongena*. *Food Chem. Toxicol.* 2009; 47: 2640-2644. DOI: 10.1016/j.fct.2009.07.026
- [13] Sudheesh S, Sandhya C, Sarah-Koshy A, Vijayalakshmi NR. Antioxidant activity of flavonoids from *Solanum melongena*. *Phytother. Res.* 1999; 13: 393-396. DOI: 10.1002/(sici)1099-1573(199908/09)13:5<393::aid-ptr474>3.0.co;2-8
- [14] Gazzani G, Papetti A, Daglia M, Berte F, Gregotti C. Protective activity of water soluble components of some common diet vegetables on rat liver microsome and the effect of thermal treatment. *Agric. Food Chem.* 1998; 46: 4123-4127. DOI: 10.1021/jf980301g
- [15] Gul S, Ahmed S, Gul H, Kaneez KF. Investigating the protective effect of *Solanum melongena*. *Asian J. Health* 2011; 1: 276-294. DOI: 10.7828/ajoh.v1i1.169
- [16] Kritchevsky D, Tepper SA, Story JA. Influence of an eggplant (*Solanum melongena*) preparation on cholesterol metabolism in rats. *Exp. Pathol.* 1975; 10: 180-183. DOI: 10.1016/s0014-4908(75)80021-4
- [17] Sudheesh S, Presannakumar G, Vijayakumar S, Vijayalakshmi NR. Hypolipidemic effect of flavonoids from *Solanum melongena*. *Plant Foods Hum. Nutr.* 1997; 51: 321-330. DOI: 10.1023/a:100.796.5927434
- [18] Shum OL, Chiu KW. Hypotensive action of *Solanum melongena* on normotensive rats. *Phytother. Res.* 1991; 5: 76-81. DOI: 10.1002/ptr.265.005.0208
- [19] Mans DRA, Toelsie J, Mohan S, Jurgens S, Muhringen M, Illes S, Bipat R. Spasmogenic effect of a *Solanum melongena* leaf extract on guinea pig tracheal chains and its possible mechanism (s). *J. Ethnopharmacol.* 2004; 95: 329-333. DOI: 10.1016/j.jep.2004.07.017
- [20] Umamageswari MS, Maniyar YA. Evaluation of anti-inflammatory activity of aqueous extract of leaves of *Solanum melongena* linn. in experimental animals. *J. Clin. Diagn. Res.* 2015; 9: 01-01. DOI: 10.7860/JCDR/2015/10777.5428
- [21] Das M, Barua N. Pharmacological activities of *Solanum melongena* Linn. (Brinjal plant). *Int. J. Green Pharm. (IJGP)* 2013; 7: 274-277.
- [22] Mohammadinejad R, Karimi S, Irvani S, Varma R.S. Plant-derived nanostructures: types and applications. *Green Chemistry.* 2016; 18:20-52. DOI: 10.1039/C5GC01403D
- [23] Somturk B, Yilmaz I, Altinkaynak C, Karatepe A, Özdemir N, Ocsoy I. Synthesis of urease hybrid nanoflowers and their enhanced catalytic properties. *Enzyme Microb. Technol.* 2016; 86: 134-142. DOI: 10.1016/j.enzmictec.2015.09.005
- [24] Akhtar MS, Panwar J, Yun YS. Biogenic synthesis of metallic nanoparticles by plant extracts. *ACS Sustain Chem. Eng.* 2013; 1: 591-602. DOI: 10.1021/sc300118u
- [25] Ankamwar B, Damle C, Ahmad A, Sastry M. Biosynthesis of gold and silver nanoparticles using *Emblica officinalis* fruit extract, their phase transfer and transmetallation in an organic solution. *J. Nanosci. Nanotechnol.* 2005; 5: 1665-1671. DOI: 10.1166/jnn.2005.184
- [26] Duman F, Ocsoy I, Kup FO. Chamomile flower extract-directed CuO nanoparticle formation for its antioxidant and DNA cleavage properties. *Mater. Sci. Eng. C* 2016; 60: 333-338. DOI: 10.1016/j.msec.2015.11.052
- [27] Demirbas A, Welt BA, Ocsoy I. Biosynthesis of red cabbage extract directed Ag NPs and their effect on the loss of antioxidant activity. *Mater. Lett.* 2016; 179: 20-23. DOI: 10.1016/j.matlet.2016.05.056
- [28] Chung JE, Tan S, Gao SJ, Yongvongsoontorn N, Kim SH, Lee JH, Ying JY. Self-assembled micellar nanocomplexes comprising green tea catechin derivatives and protein drugs for cancer therapy. *Nat. Nanotechnol.* 2014; 9: 907. DOI: 10.1038/nnano.2014.208
- [29] Altinkaynak C, Hemoglobin-metal²⁺ phosphate nanoflowers with enhanced peroxidase-like activities and their performance

- in the visual detection of hydrogen peroxide. *New J. Chem.*, 2021; 45:1573. DOI: 10.1039/D0NJ04989A
- [30] Altinkaynak C, Ildiz N, Baldemir A, Ozdemir N, Yilmaz V, Ocsoy I. Synthesis of organic-inorganic hybrid nanoflowers using *Trigonella foenum-graecum* seed extract and investigation of their anti-microbial activity. *Derim.* 2019; 3(6): 3-3. DOI: 10.16882/derim.2019.549151
- [31] Ildiz N, Baldemir A, Altinkaynak C, Özdemir N, Yilmaz V, Ocsoy I. Self-assembled snowball-like hybrid nanostructures comprising *Viburnum opulus* L. extract and metal ions for antimicrobial and catalytic applications. *Enzyme Microb. Technol.* 2017; 102: 60-66. DOI: 10.1016/j.enzmictec.2017.04.003
- [32] Baldemir A, Köse NB, Ildiz N, İlgün S, Yusufbeyoğlu S, Yilmaz V, Ocsoy I. Synthesis and characterization of green tea (*Camellia sinensis* (L.) Kuntze) extract and its major components-based nanoflowers: a new strategy to enhance antimicrobial activity. *RSC Advances.* 2017; 7(70): 44303-44308. DOI: 10.1039/C7RA07618E
- [33] Koshy DS, Das RK. Studies on the role of curcumin concentration, synthesis time, mechanism of formation, and fluorescence properties of curcumin–copper phosphate hybrid nanoflowers. *Inorg. Nano-Met. Chem.* 2020; 0: 1-8. DOI: 10.1080/24701.556.2020.1841234
- [34] Ge J, Lei J, Zare RN. Protein–inorganic hybrid nanoflowers. *Nat. Nanotechnol.* 2012; 7: 428–432. DOI: 10.1038/nnano.2012.80
- [35] Kumar D, Kumar H, Vedasiromoni JR, Pal BC. Bio – assay guided isolation of α -glucosidase inhibitory constituents from *Hibiscus mutabilis* leaves. *Phytochem. Anal.* 2012; 23: 421–425. DOI: 10.1002/pca.1375
- [36] Kumar D, Gupta N, Ghosh R, Gaonkar RH, Pal BC. α -glucosidase and α -amylase inhibitory constituent of *Carex baccans*: Bio-assay guided isolation and quantification by validated RP-HPLC-DAD. *J. Funct. Foods* 2013; 5: 211-218. DOI: 10.1016/j.jff.2012.10.007
- [37] Jeong SH, Ryu YB, Curtis-Long MJ, Ryu HW, Baek YS, Kang JE, Park KH. Tyrosinase Inhibitory Polyphenols from Roots of *Morus ihou*. *J. Agric. Food Chem.* 2009; 57: 1195-1203. DOI: 10.1021/jf8033286
- [38] Ellman GL, Courtney KD, Andres Jr V, Featherstone RM. A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochem. Pharmacol.* 1961; 7: 88-95. DOI: 10.1016/0006-2952(61)90145-9
- [39] Ozturk M, Duru ME, Kivrak Ş, Mercan-Doğan N, Türkoglu A, Özler MA. *In vitro* antioxidant, anticholinesterase and antimicrobial activity studies on three *Agaricus* species with fatty acid compositions and iron contents: A comparative study on the three most edible mushrooms. *Food Chem. Toxicol.* 2011; 49: 1353-1360. DOI: 10.1016/j.fct.2011.03.019
- [40] Lam CS, Koon HK, Chung VCH, Cheung YT. A Public survey of traditional, complementary and integrative medicine use during the COVID-19 outbreak in Hong Kong. *PLoS One* 2021; 16(7): 1-15. DOI: 10.1371/journal.pone.0253890
- [41] Kokila K, Priyadarshini SD, Sujatha V. Phytopharmacological properties of *Albizia* species: a review. *Int. J. Pharm. Pharm. Sci.* 2013; 5(3): 70-73. DOI: –
- [42] Siddiqui NA, Parvez MK, Al-Rehaily AJ, Al Dosari MS, Alam P, Shakeel F, Al Harbi HA. High-performance thin layer chromatography-based assay and stress study of a rare steroidal alkaloid solanopubamine in six species of *Solanum* grown in Saudi Arabia. *Saudi Pharm. J.* 2017; 25(2): 184-195. DOI: 10.1016/j.jsps.2016.05.003
- [43] Paoli SD, Dias AP, Capriles PV, Costa TE, Fonseca AS, Bernardo-Filho M. Effects of a tomato (*Solanum lycopersicum*) extract on the labeling of blood constituents with technetium-99m. *Rev. Bras. Farmacogn.* 2008; 18: 190-196. DOI: 10.1590/S0102-695X200.800.0200008
- [44] Asano N, Kato A, Matsui K, Watson AA, Nash RJ, Molyneux RJ, Winchester B. The effects of calystegines isolated from edible fruits and vegetables on mammalian liver glycosidases. *Glycobiology* 1997; 7: 1085-1088. DOI: 10.1093/glycob/7.8.1085
- [45] Ketprayoon T, Chaicharoenpong C. Tyrosinase inhibitory activity of some edible plants. In: Pakdibamrung K, Buaboocha T. BMB 2018. Proceedings of the international conference on biochemistry and molecular biology; 2018 Jun 20-22 Rayong, Thailand; 2018. pp.1-5.
- [46] Jo YN, Jeong HR, Jeong JH, Heo HJ. The skin protecting effects of ethanolic extracts of eggplant peels. *Korean J. Food Sci. Technol.* 2012; 44(1): 94-99. DOI: 10.9721/KJFST.2012.44.1.094
- [47] Popova I, Sell B, Pillai SS, Kuhl J, Dandurand LM. High-Performance Liquid Chromatography–Mass Spectrometry Analysis of Glycoalkaloids from Underexploited *Solanum* Species and Their Acetylcholinesterase Inhibition Activity. *Plants.* 2022; 11(3): 269-287. DOI: 10.3390/plants11030269

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